

# An Experimental Study of Microneedle-Assisted Microparticle Delivery

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Received 13 May 2013; revised 16 June 2013; accepted 21 June 2013

Published online 26 July 2013 in Wiley Online Library (wileyonlinelibrary.com). DOI 10.1002/jps.23665

**ABSTRACT:** A set of well-defined experiments has been carried out to explore whether microneedles (MNs) can enhance the penetration depths of microparticles moving at high velocity such as those expected in gene guns for delivery of gene-loaded microparticles into target tissues. These experiments are based on applying solid MNs that are used to reduce the effect of mechanical barrier function of the target so as to allow delivery of microparticles at less imposed pressure as compared with most typical gene guns. Further, a low-cost material, namely, biomedical-grade stainless steel microparticle with size ranging between 1 and 20  $\mu\text{m}$ , has been used in this study. The microparticles are compressed and bound in the form of a cylindrical pellet and mounted on a ground slide, which are then accelerated together by compressed air through a barrel. When the ground slide reaches the end of the barrel, the pellet is separated from the ground slide and is broken down into particle form by a mesh that is placed at the end of the barrel. Subsequently, these particles penetrate into the target. This paper investigates the implications of velocity of the pellet along with various other important factors that affect the particle delivery into the target. Our results suggest that the particle passage increases with an increase in pressure, mesh pore size, and decreases with increase in polyvinylpyrrolidone concentration. Most importantly, it is shown that MNs increase the penetration depths of the particles. © 2013 Wiley Periodicals, Inc. and the American Pharmacists Association *J Pharm Sci* 102:3632–3644, 2013

**Keywords:** gene gun; stainless steel; MN; microparticles; penetration depth; passage percentage; particle size; biocompatibility; biocompatibility; biomaterials; transdermal drug delivery

## INTRODUCTION

Microparticle delivery systems (e.g., gene guns) have been used for transferring genes into cells and tissues (e.g. plant tissues) for some time.<sup>1–6</sup> Typically, the operation involves a microparticle accelerator, which can deliver gene-loaded microparticles into a target (e.g., biological cells) to achieve the desired mass transfer effect (e.g., gene transfection). The PowderJect delivery system is a case in point, which has been applied to exploit the microparticle gene transfer treatment.<sup>7,8</sup> In most cases, these delivery systems are based on the principle that biocompatible microparticles loaded with genes can be accelerated to a sufficient velocity so as to penetrate the barrier function of the target tissue and thereby achieve gene delivery.<sup>9,10</sup> However, cell and tissue damages are particular problems for these microparticle delivery systems, which are discussed further later.

It is obvious from previous research on microparticle-based gene delivery that knowledge of the velocity of the microparticles and its effects on particle penetration is one of the major research points in development of these systems. A number of researchers have studied the particle velocity for various designs of gene guns. For example, Quinlan et al.<sup>11</sup> have used a conical nozzle employed at 60 bar to accelerate polymeric microparticles of 4.7, 15.5, and 26.1  $\mu\text{m}$  diameters to velocities of 350, 460, and 465 m/s, respectively. Kendall et al.<sup>12</sup> have used a converging–diverging nozzle, which has been shown to accelerate polystyrene particles of diameter 4.7  $\mu\text{m}$  to a velocity of 800 m/s at the same pressure as used by Quinlan et al.<sup>11</sup> Such developments of the delivery systems can improve the velocity of

microparticles to achieve a higher speed if compared with conical nozzles.<sup>11</sup> Mitchell et al.<sup>13</sup> have also studied the velocities of polystyrene particles (average size: 99  $\mu\text{m}$ ) for a light gas gun (LGG) proposed originally by Crozier and Hume<sup>14</sup> and gold particles (average size: 3.03  $\mu\text{m}$ ) for a contoured shock tube (CST). The particle velocity is shown to achieve 170, 250, and 330 m/s at pressure of 20, 40, and 60 bar for the LGG, respectively. The gold particles have been shown to achieve an average velocity of 550 m/s at 60 bar based on the CST. Liu et al.<sup>15</sup> have also used a CST to accelerate gold particles of diameter 2.7  $\mu\text{m}$  to a velocity of 626 m/s at 60 bar pressure. Subsequently, Liu et al.<sup>16</sup> used polystyrene particles of  $39 \pm 1$   $\mu\text{m}$  diameter to study the particle velocity for CST and found improvements relative to the LGG, which is shown to achieve a velocity of  $570 \pm 14.7$  m/s at 60 bar pressure. In recent years, Soliman et al.<sup>17</sup> have shown that a supersonic core jet can accelerate 1.8 and 5  $\mu\text{m}$  diameters gold particles to velocities of 550 and 294 m/s at 30 bar pressure. O'Brien and Lummis<sup>5</sup> have also used gold particles of core diameters 40 nm and 1  $\mu\text{m}$  to achieve maximum depths of  $31 \pm 6$  and  $50 \pm 11$   $\mu\text{m}$  in mouse ear tissue by using a Helios gene gun.

Although very high velocities of the microparticles or/and gas may seem useful in delivering the particles deep into the target tissue, they may actually damage the target from their impacts. As such, it is logical that one controls both the velocity and the mass of the microparticles and gas that impact the target. This is somewhat reflected in a study by Belyantseva<sup>18</sup> who has used a pressurized Helios gene gun to accelerate DNA-coated gold particles (1  $\mu\text{m}$  diameter) where the pressure is controlled at 14 bar. The author shows that this pressure is adequate for the penetration of the particles without excessive tissue damage. Xia et al.<sup>19</sup> have suggested that the pressure should be limited to around 14 bar to minimize damage for biolistic transfer to

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*Journal of Pharmaceutical Sciences*, Vol. 102, 3632–3644 (2013)

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soft tissue. Uchida et al.<sup>20</sup> have fired plasmid DNA into cultured mammalian cells [e.g., human embryonic kidney cell (HEK293) and human breast adenocarcinoma (MCF7) cell] using a Helios gene gun, which shows that gene transfection is achieved in these cells but the cell damage occurs if the operating pressure in the gene gun is more than 200 psi (13.78 bar). O'Brien and Lummis<sup>5</sup> have cultured HEK293 cells and used them as targets for biolistic transfection using a gene gun. This work has shown that nanoparticles can be utilized as gene carriers similar to microparticles for biolistic transfection and lessen cell damage. These researches<sup>5</sup> show that cell damage can be reduced if particle size and operation pressure are reduced as they lower the particle impact force on the cells/tissue such as those observed by Uchida et al.<sup>20</sup> In most studies, the viable dermis layer of skin is considered as the target tissue for gene-loaded microparticle delivery as the penetration depth is limited by a number of factors.<sup>11,21</sup>

In the particles delivery process, the material of the particles is also of crucial importance. In order to deliver gene-loaded particles into cells effectively, high-density materials are generally preferred as they carry a larger momentum and are expected to penetrate more into the target tissue as compared with particles of low-density materials. The most common material of the particles is gold because of its high density, low toxicity, and lack of chemical inactivity. However, gold is an expensive material. In principle, other materials such as biomedical-grade stainless steel and polystyrene may be a good replacement for gold while reducing the cost due to the lower price of these materials in comparison with gold. However, these materials have lower density compared with gold and, as such, the momentum for these microparticles would be less for the same particles size and velocity. This implies that other factor is needed to break the resistance of the target tissue for the particles to enter easily while also enhancing the penetration depths. Microneedles (MNs), which can break the resistance of the target tissue almost painlessly,<sup>22–26</sup> seem to be a promising option in this regards. However, there is little or no study at the moment that demonstrates that MNs can be useful in the delivery of dry particulates particularly at lower pressures as compared with most current gene guns, which should be operated at very high pressure.<sup>11,13,16</sup> Previously, several studies have shown that the effectiveness of the MN-based drug delivery is limited by a wide varieties of variables, for example, MN height, spaces between the needles, patch size, insertion forces, tissue characteristics such as viscoelastic properties, materials of MNs, and so on, and as such, it is necessary to choose the MNs for specific application as well as the target tissue.<sup>27–32</sup>

In addressing these points in this paper, MNs have been used to enhance the penetration depths of low-density microparticles (dry particulates) using an experimental set up that mimic particle accelerator (e.g., gene guns) in its operation principle. As model particles, we use biomedical-grade stainless steel microparticles. Further, a ground slide is used to prevent the impact of high-pressure gas on the microparticle target as discussed in more detail in the next section. The use of the ground slide gives lower particle velocities compared with the CST under the same operating conditions, which aims to reduce the cell damage. However, the purpose of the microparticle gun is to accelerate the particles to a sufficient velocity, which can penetrate into a desired depth inside the target. For a MN-based injection system, this objective could be achieved by first applying solid MNs as they help in overcoming the tissue barrier.<sup>33–35</sup>

In this study, solid MNs are used to create well-defined holes in the target, which remain open immediately after removing the MN. Hence, a number of microparticles should penetrate into the target via the holes to achieve the purpose of enhanced penetration depth. An increased penetration depth of microparticles should allow deeper tissue to be transfected if DNA/genes are coated on the microparticles. Therefore, the application of the MN-based particle delivery is a good improvement for particle injectors.

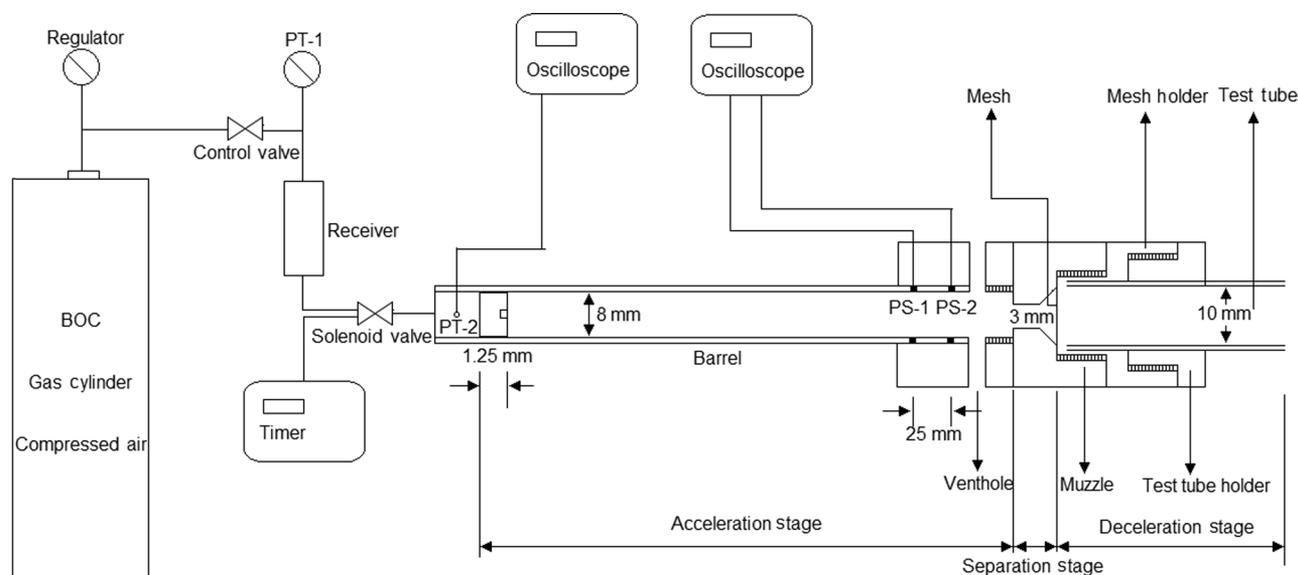
In addition to the aims discussed above, this paper aims to investigate the significance of various important factors, for example, the ground slide on the particle velocity for the MN-assisted microparticle injection. The microparticles are mixed with polyvinylpyrrolidone (PVP), compressed and bound as a cylindrical pellet for the purpose of this work. The pellet is mounted on a ground slide, which is accelerated along a barrel. The high-velocity pellet is separated by a mesh that presents a partial blockage to the flow. The work in this paper aims to determine the passage percentage and separated particle size. The paper also aims to study the effect of the MN on the microparticle penetration depth when they are fired into a homogeneous agarose gel, which is used as a model target. Agarose gel has the advantages that it can be produced with a controllable mechanical property and its transparency provides a good quality to view the microparticle penetration using optical digital microscope. In agarose gel, the microparticles follow two routes of delivery. The first route is that a number of microparticles directly penetrate into the agarose gel without going through the holes created by MNs. The second route is that the microparticles are delivered through the pierced holes created by the MNs to enhance the penetration depth inside the agarose gel. In reality, the target skin for these microparticles may be different structurally and heterogeneous, and therefore the routes of the microparticle delivery may be affected by its individual layers. However, this is not a consideration in this study as we carry out the experiments in a controlled manner using homogeneous agarose gels. The detailed information on the MN-based injection system is described in section *Experimental Design*.

## MATERIAL AND METHODOLOGY

### Materials

Biocompatible stainless steel microparticles of high sphericity equaling approximately to 0.92 were bought from LPW Technology Ltd (Daresbury, UK). Detailed characterizations of these microparticles are presented in section *Characterization of the Microparticle*. PVP purchased from Sigma–Aldrich Company Ltd. (Gillingham, UK) was dissolved in ethanol (analytical grade, 99%, obtained from Fisher Scientific Ltd., Loughborough, UK) and used to bind the microparticles to form a cohesive mixture, which could be compressed into a pellet. Agarose powder (Sigma–Aldrich Company Ltd.) was used to prepare an agarose gel, which was used a target for the microparticles penetration experiments.

Photoelectric sensors (PS) were purchased from SICK Group (Waldkirch, Germany) to detect velocity of the microparticles pellets loaded onto a ground slide. Meshes of three different pore sizes were obtained from Streme Limited (Marlow, UK). A solid MN array (Adminpatch), which has 31 needles of 1500  $\mu\text{m}$



**Figure 1.** Schematic diagram of the experimental rig.

length, was purchased from nanoBioSciences Limited Liability Company (LLC) (Sunnyvale, California).

### Experimental Design

In order to study the microparticle delivery process, an experimental rig was constructed as shown in Figure 1. Detailed information of the relevant parts of the experimental rig is listed in Table 1. The microparticle transfer process in such systems can be divided into three stages: acceleration, separation, and deceleration stage. For the acceleration stage, the pellet of microparticles attached to ground slide is accelerated to a desired speed by a pressurized gas, which was air in this study. The ground slide blocks the direct flow of gas out of the barrel and high-pressure gas is released through a venthole, avoiding impact and gas damage on the skin, agarose gel, or any other target of the microparticles. Thus, there is no gas flow in the separation and deceleration stages. In the separation stage, the pellet is released from the ground slide after hitting a stopping wall. The pellet is released from the ground slide, hits a mesh placed at the end of the barrel, which then breaks into a dispersion of microparticles by high-speed impact on an open mesh. For the deceleration stage, the separated microparticle spray forward, penetrate into the target via holes created by MNs and stop inside the target.

The detailed operating principle of the experimental rig is described as follows: a regulator is used to control the maximum gas pressure released from the gas cylinder. A control valve is located between the gas cylinder and receiver to manipulate the gas flow from the cylinder and store it in the receiver for the experiment. Additionally, a pressure transducer (PT-1) (Druck Ltd., Leicester, UK) is placed after the control valve to measure the pressure inside the receiver. A solenoid valve is used to operate the gas release from the receiver. It can open and close the gate according to a predetermined time and control the amount of gas released as required.

For the experiments, the barrel is mounted horizontally. A second pressure transducer (PT-2) (Druck Ltd.) is located at the start of the barrel to detect the driving gas pressure for accel-

erating the ground slide. This is because a large pressure drop occurs between the receiver and the barrel due to the solenoid valve and the converging section of the receiver. It means that the pressure inside the receiver is not the same as the driving pressure for ground slide acceleration. The ground slide loaded with a pellet is placed at the start of the barrel. Two PS are located at the end of the barrel, which are separated by a distance of 25 mm. They are connected to an oscilloscope to record the relevant signals and measure the velocity of the ground slide. The principle of the measurement of the ground slide speed is described in section *Experimental Data Acquisition*. In addition, two ventholes are made at the end of the barrel for release of the pressurized gas. Additionally, a mesh is placed into a muzzle at the end of the barrel and is held in place by a mesh holder. A test tube is also mounted in a holder placed at the end of the barrel to collect the separated particles and to determine the particle passage percentage (some particles remained trapped on the mesh and some rebound into the barrel). The detailed method is explained in section *Characterization of the Microparticle*. In order to investigate the effect of MN indentation on the microparticle penetration, the test tube is filled with agarose gel. The pellet is fired into this agarose gel to analyze the MN effects on particle delivery.

### Methods

#### *Experimental Data Acquisition*

*The Detection of the Ground Slide Velocity.* The velocity of the ground slide was detected by a pair of PS. The PS consist of a light source (SICK Group) and receiver (SICK Group), and they are connected to an oscilloscope to record the relevant electrical signals. Two PS were located within the barrel, which are marked as PS-1 and PS-2 in Figure 1. The barrel was made of stainless steel, and the inside surface was polished smooth to reduce friction. The space between the two PS is set at 25 mm. The working principle in this case is that the oscilloscope starts to record the signal after the ground slide reaches the position of the first sensor and covers the laser light. After the ground

**Table 1.** The Equipments and Important Parameter Values for the Experiment

Part Name	Important Variable	Material/Chemical/Other Component
Gas cylinder	Initial pressure: 200 bar Size: 146 × 23 cm <sup>2</sup> Mass: 82 kg	Compressed air Supplier: BOC (UK)
Regulator	Pressure range: 0–300 bar	Supplier: WIKA Instruments Limited (Redhill, UK)
Control valve	Pressure range: 0–100 bar	Supplier: Swagelok Company (Solon)
Pressure transducer	Range : 0–100 bar Type: XML-G100D71	Supplier: Druck Ltd. (Leicester, UK)
Receiver	Volume: 1 L	Supplier: HOKE Inc.(Spartanburg)
Solenoid valve	Pressure range: 0–100 bar	Supplier: Connexion developments Ltd. (Yate, UK)
Timer	Range: 0.1–12 s Type: H3DE-F	Supplier: OMRON Electronics Ltd (Milton Keynes, UK)
Ground slide	Diameter: 8 mm/15 mm Length: 12.5 mm	PTFE
Pellet	Diameter: 2 mm Length: 2 mm	Stainless steel microparticle
Barrel	Diameter: 8 mm/15 mm Length: 500 mm/250 mm	Stainless steel
Venthole	Diameter: 4 mm	n/a
Muzzle	Hole diameter: 3 mm	Stainless steel
Mesh holder	n/a	Stainless steel
Test tube holder	n/a	PTFE
Oscilloscope	Type: TDS 3034B	Supplier: Tektronix (Sweden, UK)
PS	Response time: 16 μs Scanning range up to 20 m Type: WLL180T	Supplier: SICK Group (Waldkirch, Germany)

slide passes the second sensor, the oscilloscope records the time for the ground slide to travel from the first sensor to the second one. Thereby, an average velocity for the speed of the ground slide was obtained based on the known distance and recorded time.

*The Analysis of the Pellet Separation.* The microparticles were compressed into the form of a cylindrical pellet for firing in the experimental rig. To make the pellet, the main materials used were biomedical-grade stainless steel particles, PVP, and 99% ethanol. At this stage of the research, no DNA or drug was loaded on the microparticles. PVP powder dissolved in ethanol was used to bind the stainless steel particles together (acting as a glue-like substance) as it has been used as a binder in many other pharmaceutical pellets.<sup>3</sup> The reason for choosing ethanol is that it helps to dry the pellet quickly due to its high volatility. The strength of the pellet is related to the PVP concentration and, as such the ethanol does not affect the binding strength of the pellet. In this study, five solutions of differing PVP concentrations were made, namely 40, 60, 75, 90, and 100 mg of PVP per mL of ethanol. On the basis of the porosity of the microparticle pellets (37.6%) and the size of the pellet, the desired amount of the PVP solution was added to 0.035 g of stainless steel powder by micropipette to fill the void space. We allow this mixture to dry for 1–2 min approximately at room temperature. When it is almost dry, we transform the powdered stainless steel with PVP solution into a solid cylindrical pellet by a pellet press (Fig. 2). As shown in the figure, the stainless steel pellet press consists of a cover, shim, main body, base, rod, and two seals. The operating procedure of this pellet press is as follows. The main body is placed on top of the base and one of the seals is inserted into the holes of the main body. Then, the powdered stainless steel microparticles containing

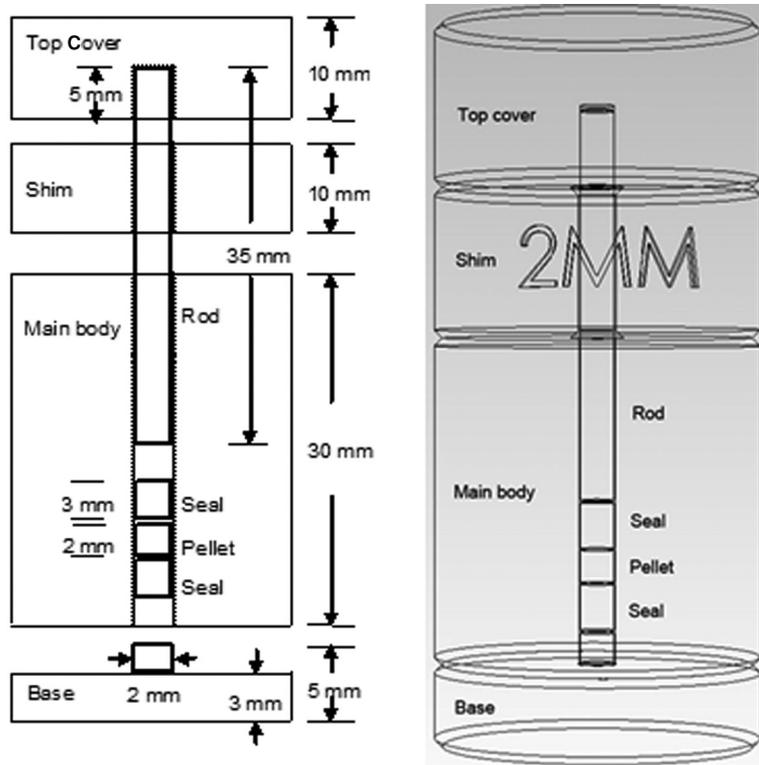
PVP solution are added followed by the second seal. The rod is placed into the cover and inserted into the main body after placing the shim on the top of main body. Finally, the top cover is pressed until there is no space between the shim and the cover. Uniformly sized pellet can be pushed out slowly using the rod directly into the ground slide to hold the pellet.

In the separation stage, the pellet is separated by a mesh and fired into an empty test tube. However, some of the separated particles are unable to pass through the mesh due to the blockage of the mesh. The analysis of the pellet separation is mainly focused on studying the passage percentage and the size of the separated particle. The mass of the pellet and test tube are measured before the experiment. The mass of the collected particles is obtained after measuring the mass of the test tube after firing. The passage percentage is calculated as:

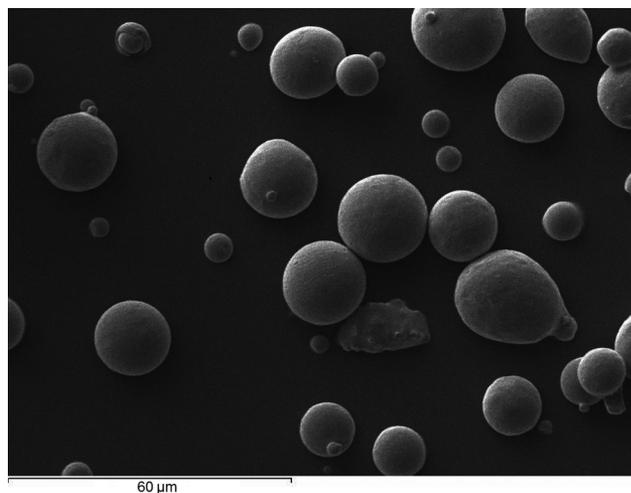
$$\text{Passage percentage} = \frac{t_1 - t_2}{m} \times 100 \quad (1)$$

where  $t_1$  is the mass of the test tube after firing and  $t_2$  is the mass of the test tube before firing,  $m$  is the initial mass of the pellet.

In addition, the separated particle sizes should be considered carefully to determine whether any large agglomerates remained, which could affect the performance of the system and penetration depth and damage to the target area. For this measurement, an adhesive-coated tape is placed at the end of the rig instead of the test tube (Fig. 1). The pellet is directly fired into the tape through the mesh, where the particles get stuck. The particle laden adhesive-coated tape is then analyzed in detail by scanning electron microscope (SEM).



**Figure 2.** Schematic diagram of the pellet press.



**Figure 3.** SEM image of the stainless steel microparticles.

### Characterization of the Microparticle

Figure 3 shows a SEM image of the stainless steel microparticles before they are made into a pellet form in this study. As can be seen, most of the particles range between diameters of 1–20  $\mu\text{m}$ , although a few larger diameter particles were found to be present. Majority of the microparticle was less than 15  $\mu\text{m}$  in diameter. From the SEM images, 30 randomly selected particles were analyzed further to calculate the average sphericity of the microparticle sample, which is found to be  $0.92 \pm 0.05$ . The actual density of the microparticles and the bulk density of

the pellet without PVP are approximately  $8 \text{ g/cm}^3$  and  $4.98 \pm 0.02 \text{ g/cm}^3$ , respectively. The porosity of the pellet without PVP is found to be  $37.6 \pm 0.3\%$  from Eq. (2)

$$\text{Porosity} = 1 - \frac{\rho_{\text{bulk}}}{\rho_{\text{particle}}} \quad (2)$$

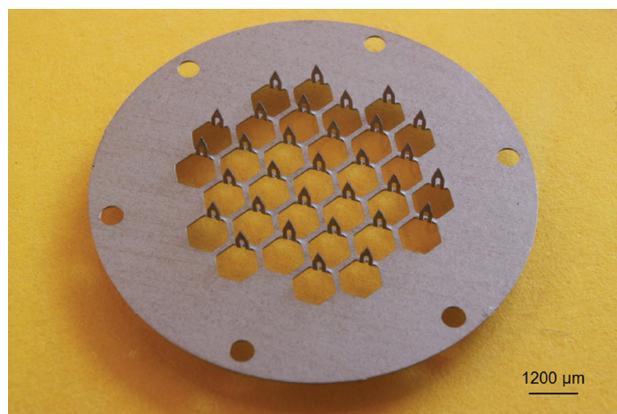
where  $\rho_{\text{bulk}}$  is the bulk density of the pellet and  $\rho_{\text{particle}}$  is the density of the particle material. In the experiment, we measure the volume of the pellet and the mass of the microparticles to obtain the bulk density, which are then used to calculate the porosity of the pellet using the equation above. The pellet has negligible amount of PVP mass and it is assumed it does not affect the bulk density or the porosity of the pellet. The ethanol that is used to dilute the PVP evaporates off from the pellet and therefore it does not affect the pellet porosity.

### Characterization of the MN

A MN patch (Adminpatch 1500), which has 31 MNs on a  $1 \text{ cm}^2$  circular patch, was used. As shown in Figure 4, the MNs are distributed as a diamond array on the patch. The space between two MNs on the side direction is  $1546 \mu\text{m}$ . The spacings between two MNs on the two diagonal directions are  $1970$  and  $3000 \mu\text{m}$ . The length, thickness, and width of each MNs are  $1500$ ,  $78$ , and  $480 \mu\text{m}$ , respectively.

### Characterization of the Mesh

Stainless steel woven meshes were chosen in this case because of their strength, higher open area, and their acceptance in pharmaceutical research. Three different mesh sizes were used in this study for pellet breakage, which are explained more in



**Figure 4.** Adminpatch 1500 microneedle array.

**Table 2.** Important Properties of the Meshes

Mesh Size	Pore Size ( $\mu\text{m}$ )	Wire Diameter ( $\mu\text{m}$ )	Open Area (%)
50	310	0.20	37
80	178	0.14	31
120	122	0.09	33

Table 2. As shown in the table, the mesh pore size decreases with an increase in mesh size. The wire diameter also has an effect on the pore size and to some extent on the fractional open area. However, for the meshes used here, the fractional open area remains approximately the same, which allows for easier comparison of the results for the different pore sizes.

## RESULTS AND DISCUSSIONS

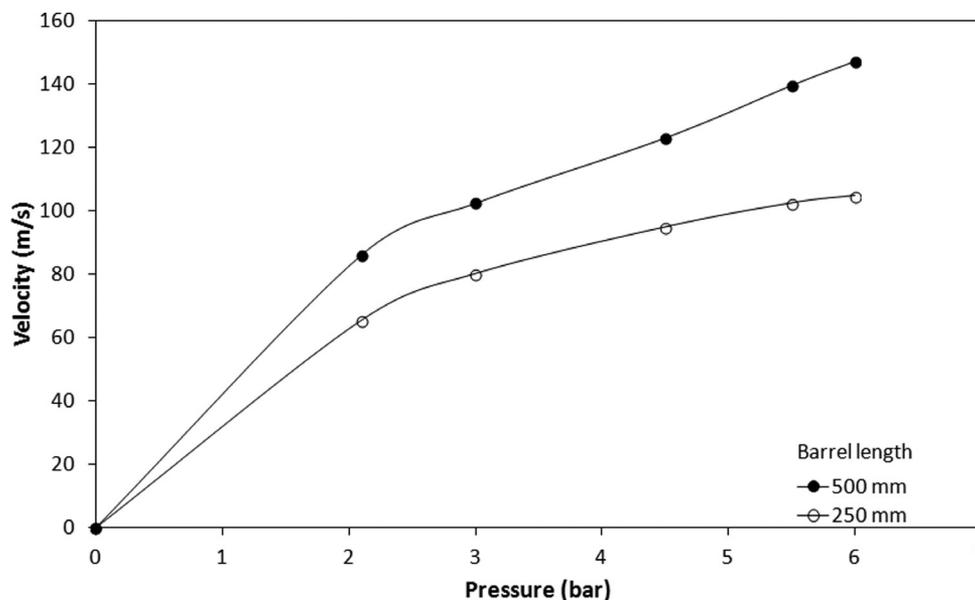
As stated earlier, the microparticles delivery process can be divided into three stages, namely, acceleration, separation, and deceleration. In the following sections, the results corresponding to each of these stages are presented and discussed.

### Particle Acceleration Stage

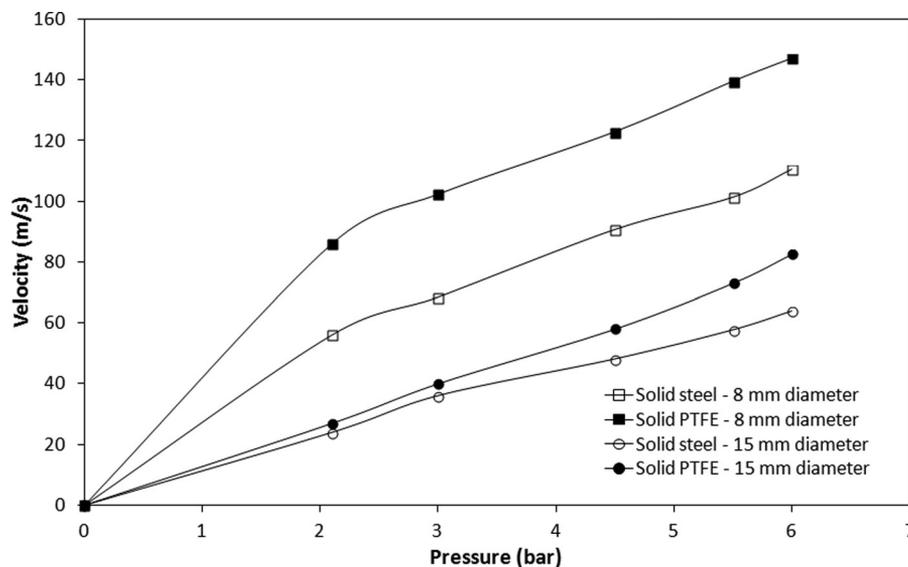
#### *The Velocity Measurement of the Ground Slide*

Some of the key variables of importance in this study are the operating pressure, the barrel diameter and length, and their effects on the velocity of the ground slide. The pellet velocity is also of importance, which is equal to the ground slide velocity at the end of the barrel. The operating pressure of the receiver (Fig. 1) is another major factor that affects the velocity of the ground slide. In the developed rig, a significant pressure drop occurs after the release of gas from the receiver due to a converging area of the receiver and losses in a solenoid valve attached to the receiver. Therefore, the pressure inside the receiver is not the actual pressure that accelerates the ground slide. The pressure at the start of the barrel is directly measured by a pressure transducer (PT-2), as explained in section *Experimental Design*. The pressure inside the gas receiver ranges from 10 to 40 bar, whereas the actual pressure to accelerate the ground slide varies between 3 and 6 bar as measured by the pressure transducer, that is, there is about 70%–85% pressure drop for the system as the gas is released from the receiver. As shown in Figure 5, the velocity of the polytetrafluoroethylene (PTFE) ground slide shows a positive correlation with the actual acceleration pressure. The particle velocity can achieve a maximum of 102, 123, 139, and 148 m/s at 3, 4.5, 5.5, and 6 bar pressures for the longest barrel. Figure 5 shows a significant difference in velocity between the two different lengths of barrel; longer barrels allow a greater time for acceleration of the ground slide. For both barrels, the velocity increases at a low rate at higher pressure. The effect of pressure on the velocity of the ground slide decreases gradually because of (1) increased friction and (2) the length of barrel is fixed, so the time for acceleration is reduced, even though the acceleration rate itself increases.

In this study, PTFE and stainless steel ground slides have been prepared to investigate how the material density affects the ground slide acceleration. The mass of the ground slide for each material made is listed in Table 3 in detail. As shown



**Figure 5.** The velocity of solid polytetrafluoroethylene ground slide against the operating pressure of gas receiver for different lengths of barrel.



**Figure 6.** The comparison of the effect of material on ground slide acceleration in the wide and narrow barrel for two different diameters of barrel/ground slide.

**Table 3.** The Key Variables Effect on the Mass of the Ground Slide

Material	Diameter (mm)	Length (mm)	Density (g/cm <sup>3</sup> )	Mass (g)
PTFE	8	12.5	2.2	1.3
PTFE	15	12.5	2.2	4.85
Stainless steel	8	12.5	8.0	4.6
Stainless steel	15	12.5	8.0	17.7

in Figure 6, the velocity of the ground slide is very different for the PTFE and stainless steel materials at the same operating condition. The density of the material affects the mass of the ground slide, making it more difficult to accelerate; hence, increased ground slide density has a negative effect on the acceleration. Similarly, Figure 6 shows that an increase of the barrel diameter causes a decrease on ground slide velocity. The mass of the ground slide increases as the barrel diameter is increased.

Overall, it is obvious that the mass of the ground slide is important in the acceleration stage. As expected, the mass increases as the barrel diameter is increased. It also increases with an increase in the material density and ground slide length. On the other hand, the velocity of the ground slide decreases with the increase in its mass. These suggest that a narrow diameter should be used for such studies as it reduces not only the ground slide diameter but also its mass.

On the basis of the above results (Figs. 5 and 6), a barrel with 8 mm diameter and 500 mm length and PTFE ground slide (12.5 mm long) were chosen for the following study on the particle separation stage. The velocity of solid PTFE ground slide is 148 m/s at 6 bar pressure. The microparticles can achieve the speed over of 600 m/s at much higher pressure for a CST, for example, at 60 bar.<sup>13,16</sup> This is because the effect of the ground slide is to reduce the effect of the fired microparticles. Mitchell et al.<sup>13</sup> have used a LGG,<sup>14</sup> which loaded the microparticles in a ground slide, so that the effect of the ground slide is investigated. They observed that the velocity of the microparticles was slow down and reduced to 170, 250, and 330 m/s at 2, 4, and 6 MPa pressure. However, the pressure drop that occurs

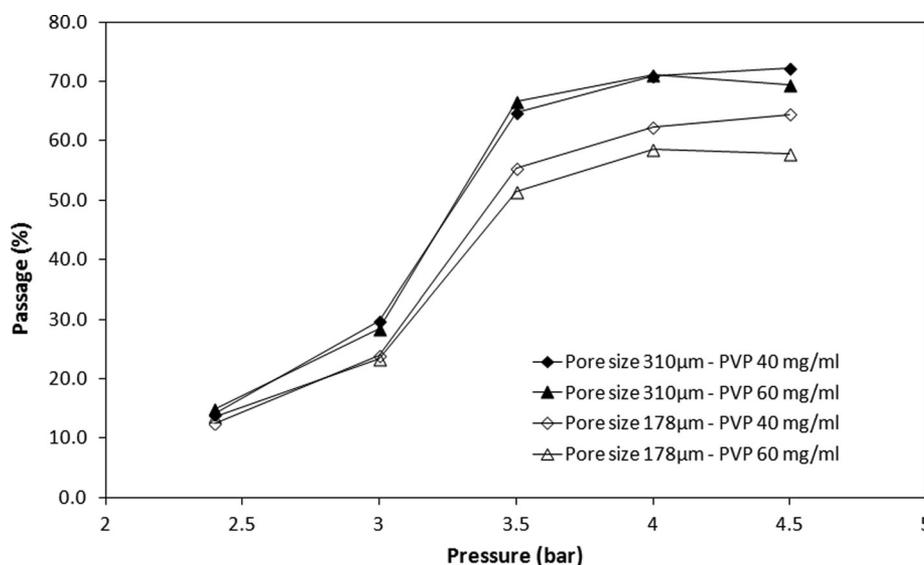
in our system means that all microparticles are accelerated to the same extent for a given barrel length. Therefore, an increased acceleration distance (barrel length) that makes up for the pressure drop effect for the current system has been chosen. Finally, the particle velocity for the current system is shown to be slightly different from that in the LGG<sup>14</sup> operated at 20 bar pressure. As a result, the particle velocity is slower if compared with the velocity obtained for other types of gene gun, largely due to the pressure drop effect. A decreased velocity decreases the microparticle penetration due to a reduction of particle momentum. However, the application of a solid MN patch has promised to remove this disadvantage as the pierced holes remain in the target tissue when the MN patch is removed and a number of microparticles can then reach further depths via the pierced holes. This is explained more in section *Deceleration Stage*.

#### Particle Separation Stage

The analysis of the pellet separation for MN-based injection system is described below. The passage percentage was analyzed in relation to the known pellet separation variables of operating pressure, PVP concentration, and mesh pore size. In addition, the microparticles size resulting from the separation stage was studied using SEM.

#### Effect of the Operation Pressure

Pellet was made using between 40 and 60 mg/mL PVP (pellet binder) concentration and results were obtained for operation between 2.4 and 4.5 bar pressures and for mesh with pore sizes of 310 and 178  $\mu\text{m}$ . As shown in Figure 7, with increasing pressure (and hence increasing velocity) the passage percentage increases rapidly at low pressures and then remains approximately constant. This is because the velocities of the pellets are larger under higher operating pressures, which cause the separated particles to gain more momentum before they are disrupted by passage through the mesh. The results show a significant increase in passage percentage from 2.4 to 3.5 bar followed by much slower increase from 3.5 to

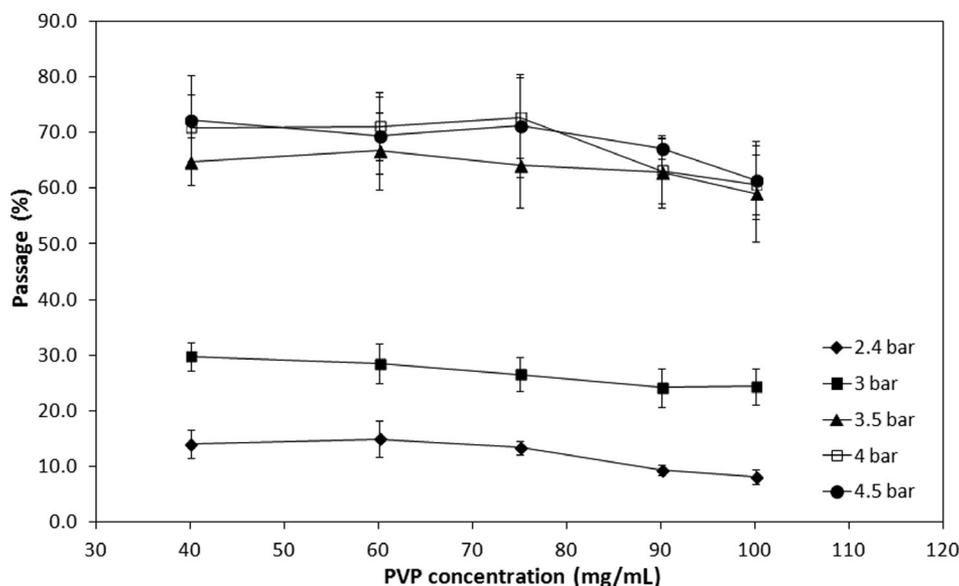


**Figure 7.** The effect of operating pressure on the passage percentage for two different PVP concentrations.

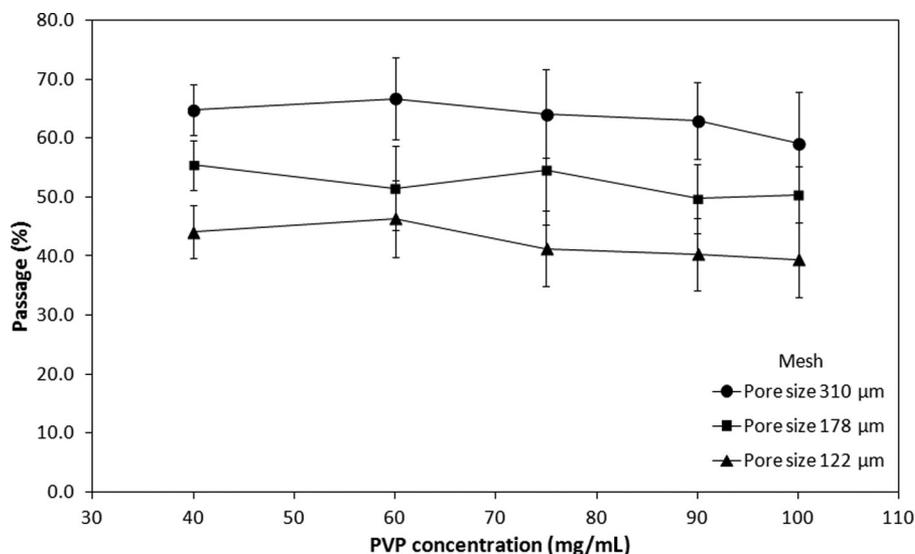
4.5 bar. This means that at lower operating pressures, there is a greater effect of pressure on the passage percentage. It is likely that the passage percentage reaches a maximum and then decreases due to some particles sticking to the mesh and some rebounding, hence not passing into the test tube (particle collector). At lower pressure conditions, the impact force on the mesh is smaller and hence the pellet is not broken up as effectively, sometime forming larger aggregates of particles, which block the mesh pores. Therefore, a larger amount of the separated particles were unable to pass through the mesh. Figure 7 also shows the passage percentage increases with an increase of mesh pore size and a decrease of PVP concentration. The detailed effects of the PVP concentration and pore size on passage percentage are explained in the following sections.

#### **Effect of PVP Concentration**

In general, the binding strength of the pellet increases with an increase in the PVP concentration, which in turn causes the passage percentage to decrease. The effect is however quite weak, as shown in Figure 8, which suggests that the passage percentage only gradually decreases with an increase of PVP concentration. The fall in passage percentage with increasing PVP concentration is due to the greater adhesive forces and increased strength of pellet, which are present in the higher PVP concentrations. The larger particles or agglomerates, that is, those that are greater in size than the mesh opening, are unable to break up as the PVP concentration increases, which means that they cannot pass through the mesh. Instinctively, the effect of PVP concentration on passage percentage should be lower at higher operating pressure. This is because the



**Figure 8.** The PVP concentration effect on the particle passage percentage at various pressures.



**Figure 9.** The particle passage percentage against PVP concentration for various mesh sizes.

impact force on the mesh is larger causing the pellet to separate more easily. However, Figure 8 shows that the range of binder concentrations used here does not lead to a significant change in the percentage of the pellet, which passes through the mesh.

#### **Effect of the Mesh Pore Size**

The effect of the mesh pore size on the pellet separation is one of the main variables that affect the particle separation. This is because the pore size is able to affect the size of the separated particles that pass through the mesh. This characteristic of the mesh could also affect the passage percentage, for example, by blocking the smallest mesh pores. To investigate the effect on the pellet separation, three different pore size meshes are studied at a constant pressure of 4.5 bar. The detailed information is explained in the Table 2, which shows that although the pore size changes, the fractional open area remains approximately constant for these meshes.

Figure 9 shows that the passage percentage exhibits significant differences for the various mesh pore sizes. As expected, the passage percentage has positive correlation with the pore size; larger pore sizes allow larger separated particles to pass through. In addition, the PVP concentration represented a negative effect on passage percentage especially for the smallest pore size mesh. The mesh with the largest pore size (about 310 μm) allowed the highest passage percentage of particles to pass through for each PVP concentration. In contrast, the mesh with the lowest pore size of 122 μm yielded the lowest passage percent. The next section discusses the effects of pore size and PVP concentration of the size distribution of the separated particles.

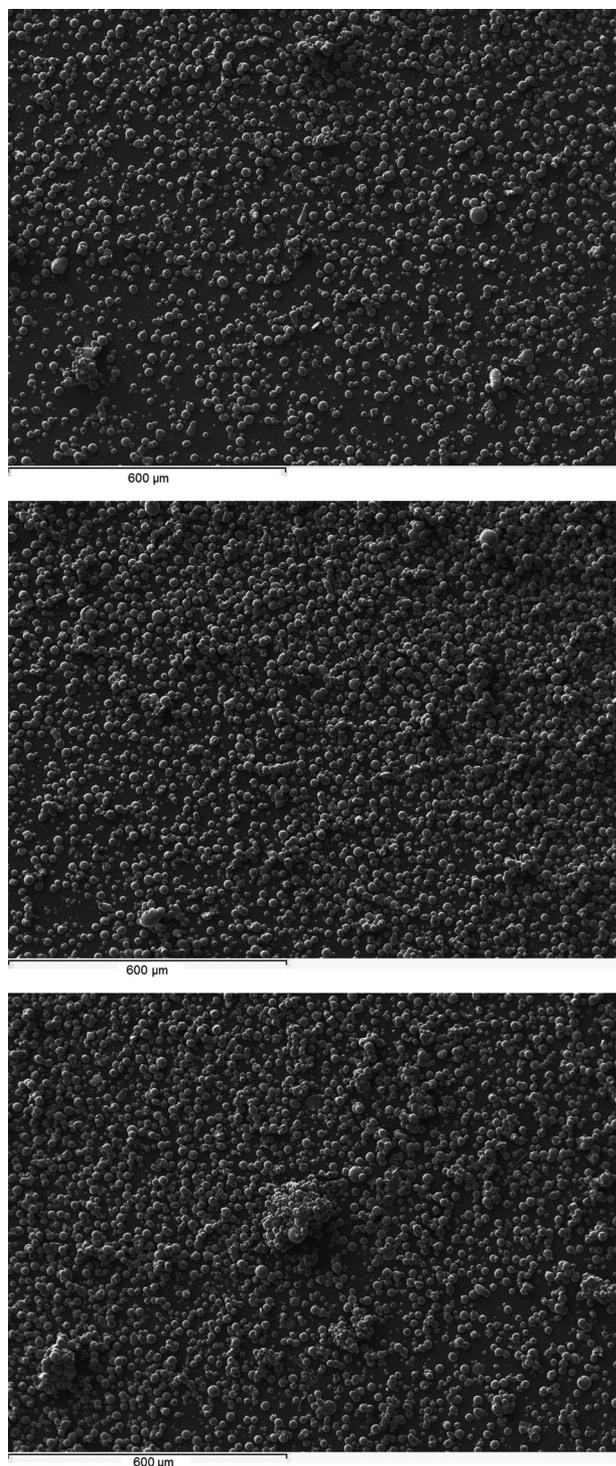
#### **The Analysis of the Separated Particle Size**

To achieve the desired particle delivery, one of the factors that are crucial to control is the size distribution of the separated microparticles. In general, the mesh pore size is able to manage the size distribution of the separated particle as it prevents passage of large particles. In this study, three different meshes were used at pressure of 4.5 bar in order to find out the effects of mesh pore size on the separated particles. The detailed information on the meshes is shown in Table 2. Figure 10a

shows an SEM of the particles produced by the mesh with pore size 122 μm; the pellet has been efficiently broken into individual particles with only a few small agglomerated particles. The maximum size of the agglomerated particle is about 50 μm. As expected, this mesh resulted in a lower passage percentage compared with the results of the other two meshes. The passage percentage improved for the mesh with pore size 178 μm (as shown in Fig. 9) and the resulting separated particles are shown in Figure 10b. This mesh also broke the pellet into individual particles and prevented passage of large sized agglomerates. The maximum size of the separated agglomerates is about 70 μm. The application of the mesh with pore size 310 μm gave the maximum passage percentage. However, as Figure 10c indicates some large agglomerated particles remain; the size of the largest agglomerate goes up to about 175 μm.

Overall, it can be concluded from this section that the size of the separated particles is controlled by the mesh pore size. The two smaller pore sizes resulted in effective pellet separation into individual particles, with relatively few large agglomerates.

The PVP concentration is also a major factor in determining the size distribution of the separated particles as it provides a binder that affects the strength of the pellet and binds the particles. Four different PVP concentrations were used to make the pellets that were fired at a pressure of 4.5 bar and mesh with pore size 178 μm. Figure 11a shows the separated particles for 40 mg/mL PVP concentration. As can be seen, the pellet was efficiently broken up into individual particles with only a few small agglomerated particles. The passage percentage decreased only slightly after increasing the PVP concentration to 60 mg/mL for the pellet and the resulting particles are shown in Figure 11b. A number of agglomerates were observed at this condition. As shown in Figure 9, the passage percentages are not significantly different for different PVP concentrations between 60 and 90 mg/mL. Figures 11b–11d show that some agglomerated particles were able to pass through pores of the 178 μm mesh. However, a 40 mg/mL PVP concentration made pellet provides a good control on the size distribution of the separated particle and a higher passage percentage, that is, it has sufficient binder strength to form the pellet that can be



**Figure 10.** SEM image of the separated particle size that is made of 40 mg/mL PVP concentration and operated at 4.5 bar pressure: (a) 122  $\mu\text{m}$  pore size (mesh size 120), (b) 178  $\mu\text{m}$  pore size (mesh 80), (c) 310  $\mu\text{m}$  pore size (mesh 50).

manipulated and mounted on the ground slide, but not so much strength that it affects particle separation.

The results show that size distribution of the separated particle is tends to be narrow for the smaller mesh pore size. Meshes with pore size 122 and 178  $\mu\text{m}$  displayed a good quality of the size distribution for 40 mg/mL PVP concentration made

of pellet. In addition, a mesh with a pore size of 178  $\mu\text{m}$  yields a higher passage percentage. Large separated particles can significantly damage the target tissue are detrimental and hence are not acceptable in this study. In addition, the strength of the pellet also does not allow the particles delivery due to a lack of particle separation. Most of the pellets should be separated into individual microparticles, and the maximum agglomerated particle size should be kept below a target of 70  $\mu\text{m}$ .

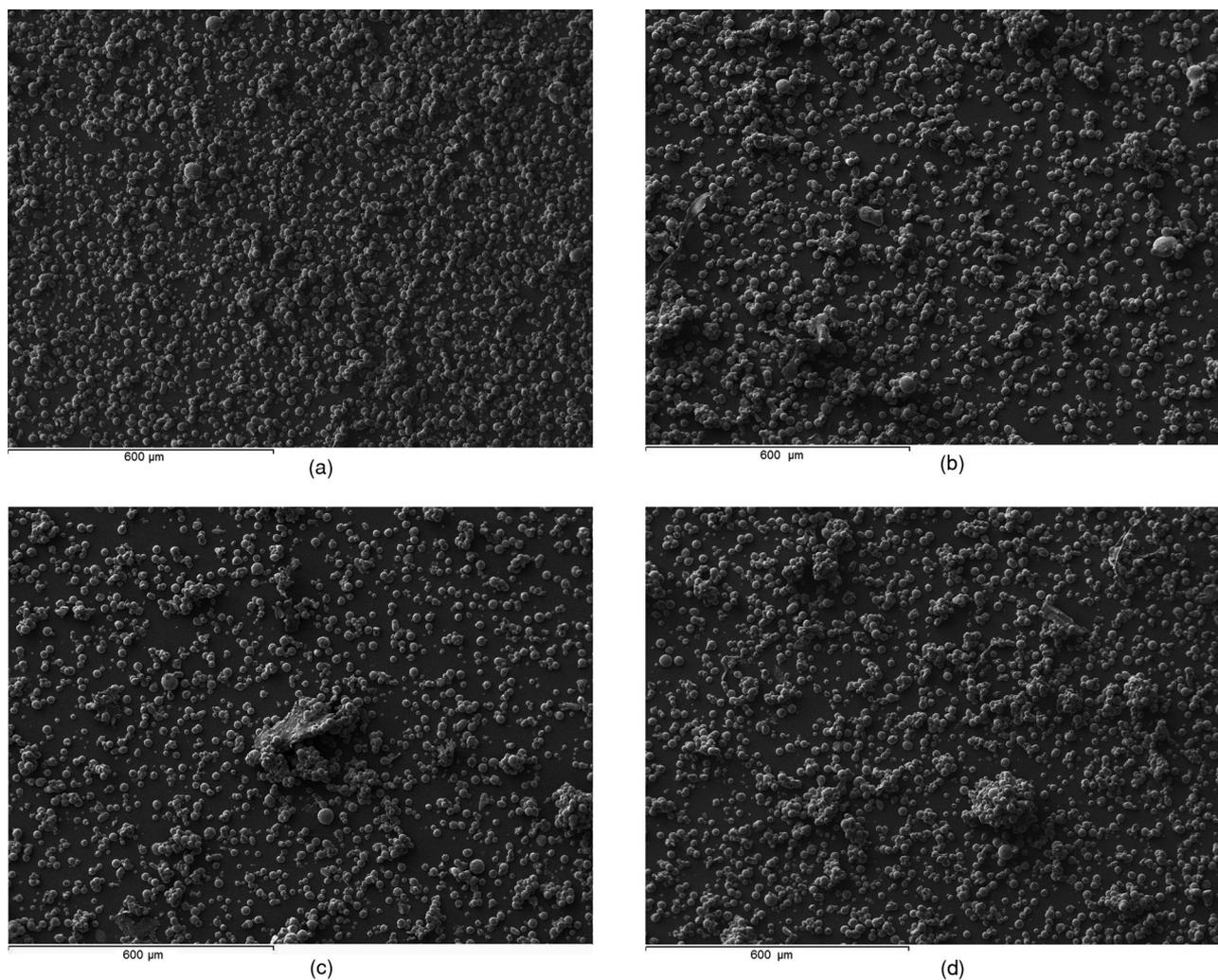
### Deceleration Stage

#### *The Microparticle Penetration in Agarose Gel*

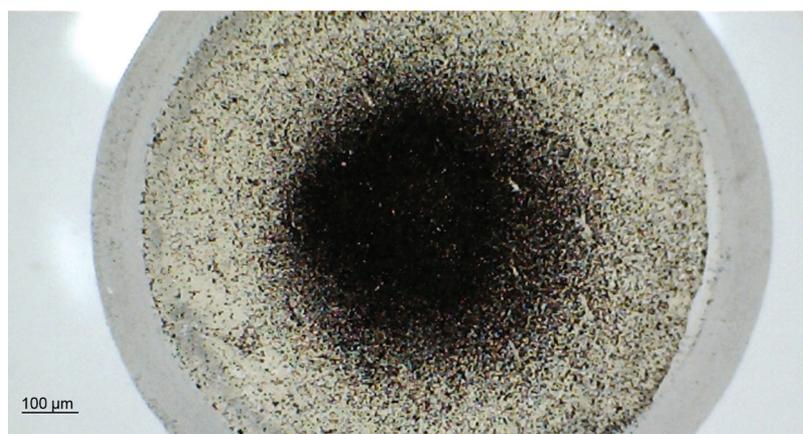
An aqueous gel made using 0.02 g/mL agarose was chosen as a target medium to study the effect of the solid MN application on the microparticle penetration. The gel is a homogenous and transparent material, which provides a good measure of the microparticle penetration. In the experiment, agarose powder is dissolved into water and heated in a microwave heater, which is then added into a sliced test tube. The test tube is covered solidly from one side with a removable film. Thus, a flat surface of agarose gel is obtained after the removal of the film when the gel is set in the test tube. The MN array is manually pressed by putting a flat plate on the back of the MN array, which provides a uniform force to pierce the MN into the gel until the backing surface of the MN just contacts the gel surface. This flat surface is used as an object of reference for the determining the insertion of the MNs. Also it is used for the measurement of the particle penetration depth. Figure 12 shows a typical distribution of the stainless steel microparticles after impact on surface of the agarose gel. As can be seen, the microparticles were nonuniformly distributed of the gel, with a maximum concentration at the center coinciding with the impact position of the pellet on the mesh. In this experiment, the MN array (see Fig. 4) had been pressed into the surface of the gel and then removed. The holes created by the MN array are clearly visible and they remain on surface of the agarose gel. The size and shape of these holes change only very slowly with time after the MNs are withdrawn. The agarose gel is a viscoelastic material like skin and the MN holes therein shrink with time. We have used 10 holes to obtain an average length of the pierced holes in the agarose gel, which is found to be approximately 720  $\mu\text{m}$  when Admipatch MN 1500 is inserted.

In order to determine the microparticle penetration, agarose gel was cut into thin slices (approximately 1 mm thick) by razor sharp blades and analyzed in more detail using a digital optical microscope (Eclipse 3100 & Digital Sight; Nikon) Kingston upon Thames, Surrey, UK. As shown in Figure 13, the holes were formed and remained in the gel after the application of the solid MN. Figure 13 also shows that the stainless steel microparticles were visible in the gel surface and within the holes. As can be seen, the microparticles seem to have a larger penetration depth compared with those that have not entered through the pierced holes on the left size of the figure. As expected, the microparticles can penetrate into a deep area via the holes. The other microparticles only have a little penetration without the hole. The maximum penetration depths are shown in Table 4 in detail.

As discussed before, Mitchell et al.<sup>13</sup> used a LGG to accelerate 99  $\mu\text{m}$  diameter polystyrene microparticles and achieved a maximum penetration depth of 150  $\mu\text{m}$  at 60 bar pressure. Kendal et al.<sup>36</sup> used a convergent–divergent device to accelerate gold particles of diameter 1.8  $\mu\text{m}$ , which achieved a maximum



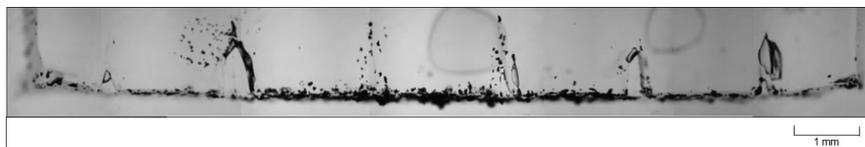
**Figure 11.** SEM image of the separated particle size that is operated at 20 bar pressure and mesh with pore size 178  $\mu\text{m}$ : (a) 40 mg/mL PVP concentration made of pellet, (b) 60 mg/mL PVP concentration made of pellet, (c) 75 mg/mL PVP concentration made of pellet, (d) 90 mg/mL PVP concentration made of pellet.



**Figure 12.** An image of the microparticle sprayed on an agarose gel.

**Table 4.** The Penetration Depths of the Microparticles

PVP Concentration (mg/mL)	Pressure (Bar)	Maximum Penetration Depth with Hole ( $\mu\text{m}$ )	Penetration Depth without Hole ( $\mu\text{m}$ )
40	4.5	$515.7 \pm 124.3$	$221.4 \pm 44.8$
	3	$508.6 \pm 137.2$	$118.7 \pm 20.3$



**Figure 13.** Optical microscope image of stainless steel microparticle penetration into agarose gel (40 mg/mL PVP, 4.5 bar, mesh with pore size 178  $\mu\text{m}$ ).

penetration depth of 78.6  $\mu\text{m}$  at 60 bar. As can be seen from Table 4, the maximum penetration depth in this study seems to improve in comparison of the results of Mitchell et al.<sup>13</sup> and Kendal et al.<sup>36</sup> However, an agarose gel of concentration 0.02 g/mL may not mimic the human skin properties exactly and as such, the implication of the mechanical properties of the target should be analyzed in more detail. Nevertheless, the results in this paper show that the application of a solid MN has a beneficial effect on the microparticle penetration depth. In addition, the agarose gel provides a good condition for the measurement of the microparticle penetration depth by a digital optical microscope. A skin mimicking agarose gel will be considered in another study to demonstrate further that MN-based system has a positive effect for microparticle delivery and the implications of the mechanical properties of the target tissue.

## CONCLUSIONS

An experimental rig involving a microparticle delivery and injection system has been built in this study to determine whether solid MNs can enhance the penetration depths of the low-density microparticles, which may be used to deliver genes and drugs. For the purpose of this design, the microparticle delivery process has been separated into three stages. For the first stage, namely, the acceleration stage, the results show that an increase in the mass of the ground slide, which carries the particles in the form of a pellet, causes a negative effect on the ground slide acceleration and hence a reduced velocity of the microcarrier pellet. The mass of the ground slide is related to its material density and size where the size is typically determined by the barrel diameter. On the basis of the present result, a narrow barrel was chosen for the study of the separation and deceleration stages as it has positive effect on the mass of the ground slide and pressure drop. For the separation stage, the passage percentage was measured using an empty test tube to collect the separated stainless steel particles, which had been broken by passage through a mesh. The results show that the passage percentage increases with pressure and mesh pore size but decreases with increasing pellet binder concentration. Increased binder concentration causes an increase of pellet strength, which seems to have a negative effect on the pellet separation. The mesh pore size affected the break up of the pellets into individual particles; larger mesh sizes allowed large agglomerated particles to pass through, which is not desirable. The mesh pore size has a significant effect on the size distribution of the separated particle and it can separate the pellet properly into individual particles. In addition, higher binder concentration pellets led to an increased number of large agglomerated particles. This is not desirable because the large particles can significantly damage the target tissue. Pellets bound with 40 mg/mL PVP yielded a higher passage percentage and a good control on the size distribution of sep-

arated particle based on the application of 178  $\mu\text{m}$  pore size mesh. For the deceleration stage, 2% concentration of agarose gel was chosen as a transparent target material to study the effect of solid MN application on microparticle delivery. The results show the pellet is well separated and sprayed onto the target; a number of stainless steel microparticles can penetrate a deep area inside the gel due to the holes created by the solid MN application. The maximum penetration depth is comparable with previous study<sup>13,36</sup> and in some cases shows a significant improvement, but without the need for high-pressure gas flows, which can damage soft tissues. However, this should be investigated further in a future study.

## ACKNOWLEDGMENTS

Loughborough University (UK) is acknowledged for providing a PhD studentship to Dongwei Zhang, which made this work possible. Further, the technical supports from Mr. Tony Eyre, Mr. Mark Barron, Mr. Jim Muddimer, Mr. Terry Neale, and Mr. Steve Bowler are gratefully acknowledged.

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